

Production of ^{88}Y for gamma-ray emission imaging

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Radioimmunotherapy (RIT) is an internal radiation therapy that uses radiolabeled drugs, in particular particularly in monoclonal antibodies (mAbs) or peptides. ^{90}Y emits highly cytotoxic β^- ray and is thus a promising radionuclide for use in RIT. However, ^{90}Y cannot be readily imaged by nuclear medicine imaging modalities, because ^{90}Y is a pure β^- emitter.¹⁾ On the other hand, ^{86}Y emits β^+ rays, which can be detected by PET.¹⁻³⁾ In addition, ^{86}Y -labeled drugs (mAbs or peptides) display identical biodistributions to ^{90}Y -labeled drugs because ^{86}Y is chemically identical to ^{90}Y .¹⁾ Therefore, in recent years, ^{86}Y has attracted attention as an attractive surrogate for studying ^{90}Y -labeled drugs. However, the physical half-life of ^{86}Y ($T_{1/2} = 14.7$ h) is shorter compared to that of ^{90}Y ($T_{1/2} = 64.1$ h), and thus, it is not suitable as a surrogate for investigating serial biodistribution of RIT drugs with long biological half-lives, such as mAb, which remain circulating in vivo for weeks.⁴⁾ A chemically identical surrogate with a longer half-life is desirable for development phases of ^{90}Y -labeled drugs.

^{88}Y is chemically identical to ^{90}Y and has a long half-life of $T_{1/2} = 106.6$ d. Moreover, ^{88}Y emits γ rays with energies of 898 and 1836 keV, which can be detected using semiconductor Compton cameras through gamma-ray emission imaging (GREI).⁵⁾ Therefore, the imaging of ^{88}Y -labeled drugs with GREI has the potential ability to investigate the serial biodistribution of ^{90}Y -labeled drugs with a long biological half-life, in particular, in preclinical studies. The final purpose of our study is to develop an imaging method for ^{88}Y -labeled drugs through GREI. In this study, we produced ^{88}Y for the GREI experiment.

^{88}Y was produced by the $^{nat}\text{Sr}(d,x)^{88}\text{Y}$ reactions. To prepare a ^{nat}SrO pellet target with a diameter of 10 mm, approximately 400 mg of $^{nat}\text{SrCO}_3$ (Wako Pure Chemical Industries, Ltd., chemical purity: 99.99%) was heated for 2 h at 1000°C and pressed at 1.6 t. The pellet was covered with a 10- μm Al foil (chemical purity: 99.999%). The target was irradiated with a 24-MeV deuteron beam supplied from the RIKEN AVF cyclotron. The irradiation was performed for 5 h at a beam current of approximately 1.5 particle μA .

Thirty-nine days after the irradiations, ^{88}Y was chemically isolated from the ^{nat}SrO target by extraction chromatography using Ln-resin (Eichrom Technologies, Inc., particle size: 50-100 μm) filled in a Muromac column (Muromachi Technos Co., Ltd., internal diameter: 5 mm, height: 50 mm). The Ln-resin column was washed in

advance with 3 mL of water, 10 mL of 10 M HNO_3 , and then 4 mL of water, and was pre-equilibrated with 2 mL of 1 M HNO_3 . The irradiated ^{nat}SrO target was dissolved in 1 M HCl and evaporated to dryness on a hot plate and under a heat lamp. The residue was dissolved in 5 mL of 1 M HNO_3 , and was evaporated to dryness. Subsequently, the residue was dissolved in 5 mL of water and 2 mL of 1 M HNO_3 , and was evaporated to dryness. Then, the residue was dissolved in 2 mL of 1 M HNO_3 and loaded onto the Ln-resin column. The resin was then washed with 16 mL of 1 M HNO_3 . ^{88}Y was eluted from the resin with 10 mL of 10 M HNO_3 . The eluted solution was heated to dryness, and 2 M HCl was added to the residue.

The γ -ray spectrum of the final purified product is shown in Fig. 1. Approximately 10 MBq of ^{88}Y was obtained. The radiochemical yield of ^{88}Y in the chemical isolation process was approximately 80%. In the next experiment, we plan to synthesize ^{88}Y -labeled drugs and try to visualize their biodistribution using GREI.

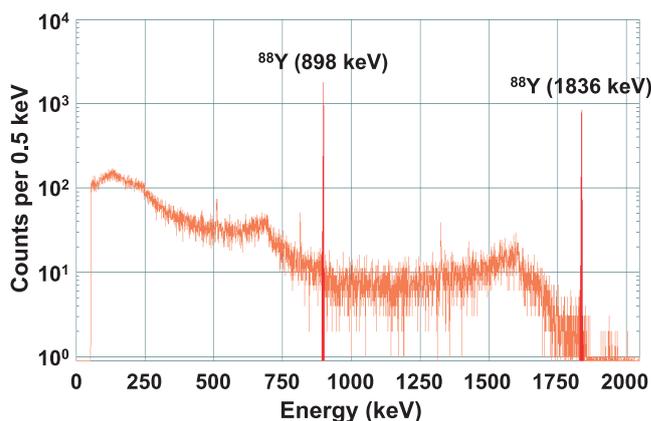


Fig. 1. Gamma-ray spectrum of ^{88}Y after the chemical isolation.

References

- 1) T. K. Nayak et al.: *Med. Chem.* **7**(5), 380 (2011).
- 2) S. Palm et al.: *J. Nucl. Med.* **44**(7), 1148 (2003).
- 3) T. K. Nayak et al.: *Eur. J. Nucl. Med. Mol. Imaging.* **37**, 1368 (2010).
- 4) D. R. Mould et al.: *Curr. Opin. Drug Discov. Devel.* **10**(1), 84 (2007).
- 5) S. Motomura et al.: *J. Anal. At. Spectrom.* **23**, 1089 (2008).

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